

# Simultaneous Estimation of Minoxidil and Finasteride by RP-HPLC in Presence of Soy Lecithin Excipient in Lotion Dosage Form

I.Ponnilarasan\*, Mansoor.K.P, K.K.Sivakumar

KMCH College of Pharmacy, Kovai Estate, Kalapatty Road, Coimbatore, Tamil Nadu India.

\*Corresponding Author

**Abstract:** A simple, fast, precise, selective and accurate RP-HPLC method was developed and validated for the simultaneous determination of Minoxidil and Finasteride from bulk and liquid formulations in the presence of soy lecithin excipient. Chromatographic separation was achieved isocratically on a Phenomenex C18 column (250×4.6 mm, 5 μ particle size) using a mobile phase, Phosphate buffer: acetonitrile: methanol (50:50 v/v) (adjusted to pH 5.8 with NaOH) in the ratio of 30:70. The flow rate was 1.2 ml/min and effluent was detected at 211nm. The retention time of Minoxidil and Finasteride were 3.16, and 8.23 min respectively. Linearity was observed in the concentration range of 50 to 250 μg/ml, 1 to 5 μg/ml for Minoxidil and finasteride with correlation coefficient 0.999 for both the drugs. Percent recoveries obtained for both the drugs were 99.275-99.72 and 98.54-99.3% w/w. The method was validated according to the ICH guidelines with respect to specificity, linearity, accuracy, precision and robustness. The method developed can be used for the routine analysis of Minoxidil and Finasteride in combined dosage form.

**Key words:** RP-HPLC Method; Phenomenex; Minoxidil; Finasteride; soy lecithin

## I. INTRODUCTION

Finasteride, N-(1,1-dimethylethyl)-3-oxo-(5α,17β)-4-azaandrost-1-ene-17-carboxamide, a type II 5 alpha reductase inhibitor, The mechanism of action of Finasteride is based on its preferential inhibition of Type II 5a-reductase through the formation of a stable complex with the enzyme. Inhibition of Type II 5a-reductase blocks the peripheral conversion of testosterone to DHT, resulting in significant decreases in serum and tissue DHT concentrations, minimal to moderate increase in serum testosterone concentrations, and substantial increases in prostatic testosterone concentrations. As DHT appears to be the principal androgen responsible for stimulation of prostatic growth, a decrease in DHT concentrations will result in a decrease in prostatic volume (approximately 20-30% after 6-24 months of continued therapy). In men with androgenic alopecia, the mechanism of action has not been fully determined, but finasteride has shown to decrease scalp DHT concentration to the levels found in hairy scalp, reduce serum DHT, increase hair re growth, and slow hair loss, Minoxidil, 6-piperidin-1-ylpyrimidine-2,4-diamine 3-oxide. The mechanism by which

minoxidil promotes hair growth is not fully understood. Minoxidil contains the nitric oxide chemical moiety and may act as a nitric oxide agonist. Similarly, minoxidil is a potassium channel opener, causing hyper polarization of cell membranes. Minoxidil is less effective when there is a large area of hair loss. In addition, its effectiveness has largely been demonstrated in younger men who have experienced hair loss for less than 5 years. Minoxidil use is indicated for central (vertex) hair loss only. Minoxidil is also a vasodilator. Hypothetically, by widening blood vessels and opening potassium channels, it allows more oxygen, blood, and nutrients to the follicle. This may cause follicles in the telogen phase to shed, which are then replaced by thicker hairs in a new anagen phase.

Literature review reveals various works on UV<sup>[1-4]</sup>, HPLC<sup>[4-7]</sup>, of Minoxidil, UV<sup>[8]</sup>, HPLC<sup>[9-12]</sup>, LC MS/MS<sup>[13]</sup>, UPLC<sup>[14]</sup>, UPLC MS/MS<sup>[15]</sup> of Finasteride. To the author's best knowledge, no method has been reported for simultaneous estimation of Minoxidil and Finasteride in combined lotion dosage form. Hence, it was thought to be of interest to develop RP-HPLC methods for simultaneous estimation of Minoxidil and Finasteride in bulk as well as in combined lotion dosage form.

## II. EXPERIMENTAL

### Chemicals, Reagents and Solutions

Minoxidil (>98%) was purchased from Cipla R&D Mumbai, Finasteride purchased from Dr.Reddys Hyderabad. Acetonitrile and methanol (HPLC-grade) was purchased from Sigma Aldrich. Analytical grade potassium dihydrogen phosphate was obtained from SD Fine chemicals LTD Mumbai. The other chemicals were of analytical-reagent grade. Morr F (Batch No. TIM 12002) were manufactured by Intas Pharmaceutical Industry Co., Ltd, Ahmedabad. An amount of minoxidil and finasteride was weighed accurately, dissolved in methanol and then diluted to 1mg/ml. Working standard solutions were prepared by diluting the stock solution with the mobile phase. All standard solutions were stored at 20°C. Before RP-HPLC analysis, the working solution was filtered through a 0.22-μm membrane prior to injection into the HPLC system.

### Sample Preparation

One gram liquid was extracted with 10ml of methanol and filtered through whatmann filter paper No: 41. The final mixed sample solution was prepared, corresponding to 150ug/ml of Minoxidil and 3µg/ml of finasteride. After filtration through a 0.22-µm membrane filter, the chromatographic analysis was carried out.

### Chromatography

Chromatographic separation and quantitative determination were performed using a high-performance liquid chromatography containing a Shimadzu pump (model LC-20AT), a PDA detector (*SPD-6AV*, Shimadzu), a Rheodyne 7120 with a 20-µL loop. As the stationary phase, a Phenomenex RP-18 column, 5 µm particle size, 250 × 4 mm (*Merck, Darmstadt, Germany*), was used. The mobile phase consisted of 70 vol of acetonitrile: methanol and 30 vol of phosphate buffer (pH 5.8). The flow rate of the mobile phase was 1.2 ml/min. The wavelength of the PDA detector was set at 211nm.

### Method Validation

A linear calibration curve was generated by plotting the peak area versus minoxidil and finasteride concentrations. The slope, intercept, and correlation coefficient values were estimated by using a least squares regression analysis. The limit of detection (LOD) was defined as 3 times the signal-to-noise ratio (S/N). The limit of quantification (LOQ) was defined as the lowest concentration in the linear calibration curve. Validation of HPLC method for minoxidil and finasteride was then carried out. Quality control samples containing 150ug/ml of minoxidil and 3ug/ml of finasteride were used to evaluate the precision and accuracy of the proposed method. Intraday variability and precision were determined by analyzing the quality control sample in duplicate on the same day. Interday variability and precision were evaluated by injecting duplicate processed samples at each control concentration for 5 days. The assay precision was reflected by the relative standard deviation (RSD). The accuracy of the method was assessed by the use of the standard addition technique. Known amounts of pure minoxidil and finasteride were added to the liquid dosage form in which the sample concentration was determined. The amounts of analytes recovered were estimated by use of the regression equation of the calibration plots. The relative recovery in the liquid dosage form was evaluated at 50ug/ml of minoxidil and 1ug/ml of finasteride. The accuracy study was performed three times.

## III. RESULTS AND DISCUSSION

### Sample Analysis

The developed RP-HPLC method was applied to analyze Minoxidil and finasteride in Morr F liquid dosage form. The chromatogram of a sample is shown in Fig. 3. Based on the peak area of Fig. 3, the content of Minoxidil and finasteride in

Morr F liquid dosage form is about 50mg/ml and 1mg/ml with an RSD of 0.91% (n= 6) according to the previously established linear regression equation.

### Linearity

Under the above described experimental conditions, a linear relationship was established by plotting the peak area against the drug concentration. The concentration ranges were found to be 50–250ug/ml and 1–5ug/ml for minoxidil and finasteride respectively. Linear regression analysis of the data gave the following equations:

Write  $y=mx+c$  equation of both drugs

Statistical analysis of the data gave a high value of the correlation coefficient (r) of the regression equation; small values of the intercept (Sa) and of slope (Sb) and small values of the percentage relative standard deviation (Table No 1). These data proved the linearity of the calibration graph.

### Limit of Quantitation (LOQ) and Limit of Detection (LOD)

LOQ and LOD were calculated according to ICH Q2R1 recommendations using the following equation [26]:

$$LOQ = 10XSD/Slope \text{ and } LOD = 3.3 XSD/Slope$$

Where SD is the standard deviation of the intercept of the calibration curve

LOQ values were found to be 18.11ug/ml and 2.54ug/ml, while LOD values were found to be 5.97ug/ml and 0.83ug/ml for minoxidil and finasteride respectively.

### Accuracy and Precision

In order to further evaluate the validity of the proposed method for the assay of minoxidil and finasteride in real samples, a recovery experiment was carried out. The Sample was spiked with a known amount of pure drug and the recovery was determined. The satisfactory recovery of 99.10-99.72 for minoxidil and 98.54-99.34 for finasteride in the Morr F liquid dosage form indicates that the proposed method is reliable for the quantification of the Minoxidil and finasteride content in the sample.

Intraday and interday precisions were assessed using single concentration and six replicates. The relative standard deviations were found to be very small, indicating reasonable repeatability and intermediate precision of the proposed method (**Table no 2**)

Precision data for the determination of Minoxidil and finasteride by the proposed method

### Robustness of the Method

The stability of the peak area with deliberate changes in the experimental parameters indicates the robustness of the proposed method. These parameters include methanol: ACN concentration ( $70 \pm 1\%$  (v/v)) and the ionic strength of buffer ( $0.04 M \pm 0.005$ ). These minor changes did not greatly affect the peak area ratios of any of the drugs.

### Selectivity

The selectivity of the method was investigated by observing any interference encountered from common tablet excipients. It was shown that these compounds did not interfere with the results of the proposed method.

### IV. CONCLUSION

It was concluded that the developed RP-HPLC method was found to be very simple, reliable and selective for providing satisfactory accuracy and precision. The methods are suitable for routine quantitative analysis in pharmaceutical dosage forms. In this present study the retention time of minoxidil and finasteride was found to be 3.16min and 8.23min respectively. Furthermore the method has been shown to be specific and selective. From this current study concluded that lecithin in the formulation as the excipient used for permeation enhancer is which has strong absorbance in the UV region it is minimized by this extraction method methanol and the mobile phase used for the RP-HPLC method developed for drugs.

### BIBLIOGRAPHY

- [1]. Zahid A Zaheer, Shahed Mirza, Ismail Moazzam and Imran W Sayad, 2012, UV-Spectrophotometric determination of minoxidil and its application to the assay in pharmaceutical dosage forms, *Der Pharma Chemica*, 2012, 4 (1), Pg no 568-573.
- [2]. Hemanth K Gaidhane, Jagdish P Bidada, Akshay S Bhusari, Manjusha S Navkhare, Ganesh P Diwanka, Ashish H Tiwari. Development and Validation of Stability Indicating HPLC Method for the estimation of Minoxidil and related substance in topical formulation, *Journal of Pharmacy Research*, 2011: 4(12), Pg no: 4481-4484.
- [3]. Maryam Bordbar, Ali Yeganeh-Faal, Jahanbakhsh Ghasemi, Mohammad Mahdi Ahari-Mostafavi, Nahid Sarlak and Mohammad Taghi Baharifar. Simultaneous spectrophotometric determination of minoxidil and tretinoin by the H-point standard addition method and partial least squares, *Chemical Papers*, 2009: 63(3), Pg no: 336-344
- [4]. Bonazzi D, Di Pietra AM, Gatti R, Cavrini V. Determination of minoxidil in pharmaceutical formulations by difference spectrophotometry and liquid chromatography (HPLC), *Farmaco Societa chimica italiana*. 1990, 45(6): Pg no: 727-735.
- [5]. Carrum G, Abernethy DR, Sadhukhan M, Wright CE 3rd. Minoxidil analysis in human plasma using high-performance liquid chromatography with electrochemical detection. Application to pharmacokinetic studies. *Journal of Chromatography*. 1986 Aug 22; 381(1):127-35
- [6]. Zarghi A. Shafaati, S.M. Foroutan A. Khoddam, Rapid determination of minoxidil in human plasma using ion-pair HPLC, *Journal of Pharmaceutical and Biomedical Analysis*, Volume 36, Issue 2, 29 October 2004, Pg no 377-379
- [7]. Jeffrey Hurst.W, Robert A. Martin. The analysis of phospholipids in soy lecithin by HPLC, *Journal of the American oil chemists' society*, Volume 61, Number 9 (1984), Pg no.1462-1463.
- [8]. Manish Kumar Thimmaraju, Venkat Rao, Srikanth Gurralla, Jayapal Reddy. UV Spectrophotometric Method for Simultaneous Determination of Finasteride and Tamsulosin in combined dosage form, *International Journal of Pharmacy and Biological Sciences*, 2011: 1(3), Pg no: 303-310
- [9]. Basavaiah.K, Somashekar.B.C, Determination of Finasteride in Tablets by High Performance Liquid Chromatography, *E-Journal of Chemistry*, 2006: 4, (1), Pg no: 109-116
- [10]. Dipti B. Patel, Natubhai J. Patel. Validated RP-HPLC and TLC methods for simultaneous estimation of tamsulosin hydrochloride and finasteride in combined dosage forms, *Acta Pharmaceutica*, 2010: 60, Pg no: 197-205
- [11]. Srinivas.G, KishoreKumar.K, Yarram Rama Koti Reddy, Mukkanti.K, Gangaram V. Kanumula, Madhavan.P, Validated stability indicating LC method of assay and related substances for Finasteride, *Journal of Chemical and Pharmaceutical Research*, 2011: 3(6), 987-996.
- [12]. Akheel.A.Syed, Mungalimane K., Amshumali. LC determination of finasteride and its application to storage stability studies, *Journal of Pharmaceutical and Biomedical Analysis* 25 (2001) 1015-1019.
- [13]. Xiaohong Chen, Erin R. Gardner, Douglas K. Price, and William D. Figg. Development and Validation of an LC-MS Assay for Finasteride and its Application to Prostate Cancer Prevention Trial Sample Analysis, *Journal of Chromatographic Science*, 2008: 46, 356-361.
- [14]. Szabolcs Fekete, Jenő Fekete, Katalin Ganzler. Validated UPLC method for the fast and sensitive determination of steroid residues in support of cleaning validation in formulation area, *Journal of Pharmaceutical and Biomedical Analysis* 49, 2009, 833-838.
- [15]. Phapale PB, Lee HW, Lim MS, Kim EH, Kim SD, Park J, Lee M, Hwang SK, Yoon YR. Rapid determination of finasteride in human plasma by UPLC-MS/MS and its application to clinical pharmacokinetic study, *Journal of Chromatography. B. Analytical Technologies in the Biomedical and Life Sciences*, 2010 Jun 15; 878(20):1718-23.

Table No 1: Analytical performance data

Parameter	Minoxidil	Finasteride
Linearity range ( $\mu\text{g mL}^{-1}$ )	50-250	1-5
Intercept (a)	95871	14750
Slope (b)	43911	38369
Correlation coefficient (r)	0.999	0.999
Percentage relative standard deviation, %RSD	1.303	0.816
Limit of detection, LOD ( $\mu\text{g/ml}$ )	5.97	0.83
Limit of quantitation, LOQ ( $\mu\text{g/ml}$ )	18.11	2.54

Table No 2: Assay of the drugs in pure form

Compound	Amount taken	Amount recovered	%recovery
Minoxidil	80	79.42	99.275 $\pm$ 0.43
	100	99.1	99.10 $\pm$ 0.40
	120	119.67	99.72 $\pm$ 0.27
Mean $\pm$ SD		99.39 $\pm$ 20.19	
Finasteride	80	78.83	98.54 $\pm$ 0.45
	100	99.10	99.10 $\pm$ 0.40
	120	119.21	99.34 $\pm$ 0.37
Mean $\pm$ SD		99.04 $\pm$ 20.1	

Table No 3: Inter day studies

Day	Conc. of MINOX ( $\mu\text{g/ml}$ )	Peak Area	% RSD	Conc. of FNS ( $\mu\text{g/ml}$ )	Peak Area	% RSD
Day 1	150	6480024	0.65	3	1285736	1.04
		6553981			1279543	
		6467521			1288757	
		6538754			1268774	
		6512345			1254718	
		6577874			1287169	
Day 2	150	6478956	0.85	3	1276548	1.25
		6533789			1286738	
		6536578			1297893	
		6578893			1287945	
		6576892			1250870	
		6437789			1278754	
Day 3	150	6475022	0.57	3	1277691	0.87
		6556979			1285367	
		6499739			1274705	
		6566179			1293903	
		6489911			1265241	
		6498986			1291547	

Table No 4: Intraday studies

No of Injection	Conc. Of MINOX (µg/ml)	Peak Area	% RSD	Conc. of FNS (µg/ml)	Peak Area	% RSD
6	150	6549350	1.05132	3	1201452	1.3937
		6481452			1216530	
		6479201			1194672	
		6538641			1214525	
		6474394			1214525	
		6357835			1201962	

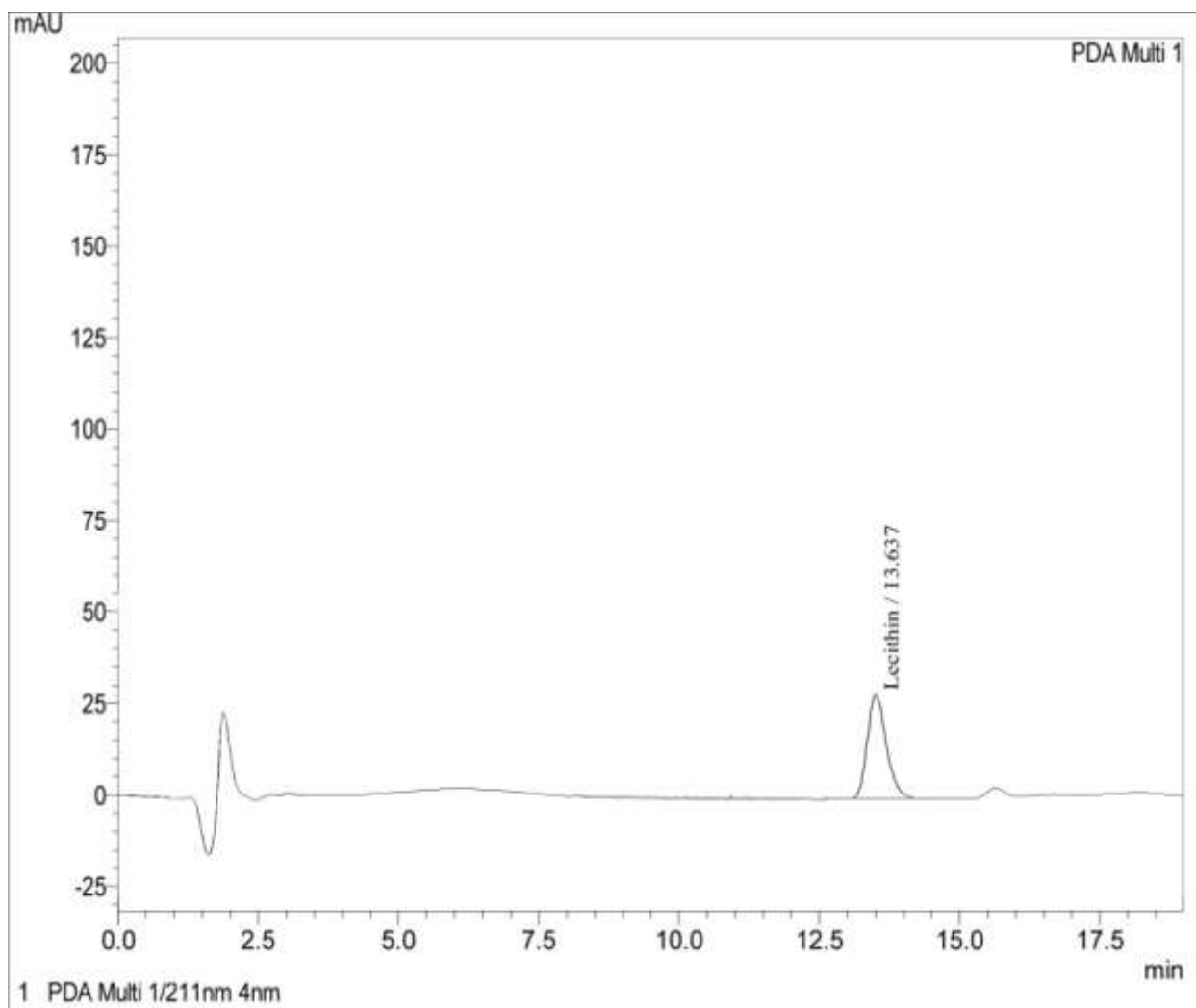


Figure 1. Standard chromatogram of Lecithin

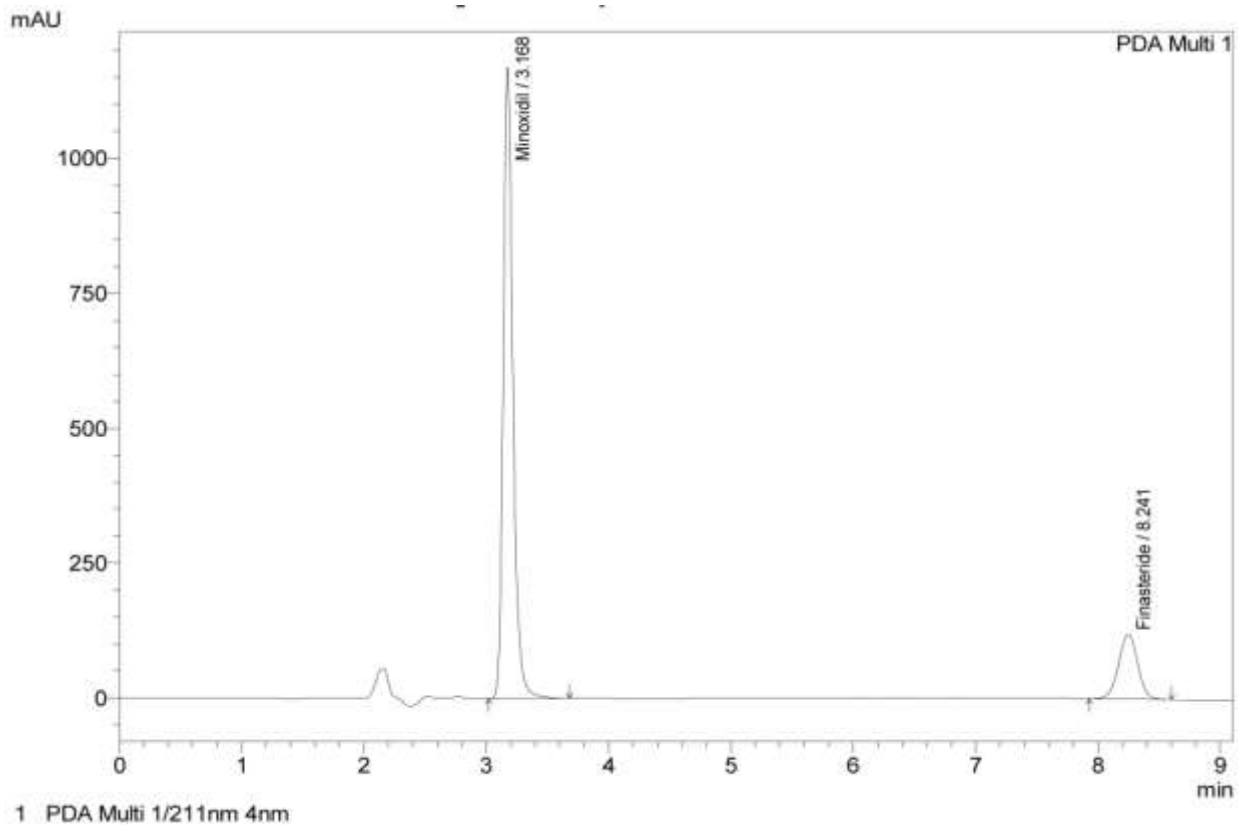


Fig 2. Standard chromatogram of Minoxidil and Finasteride

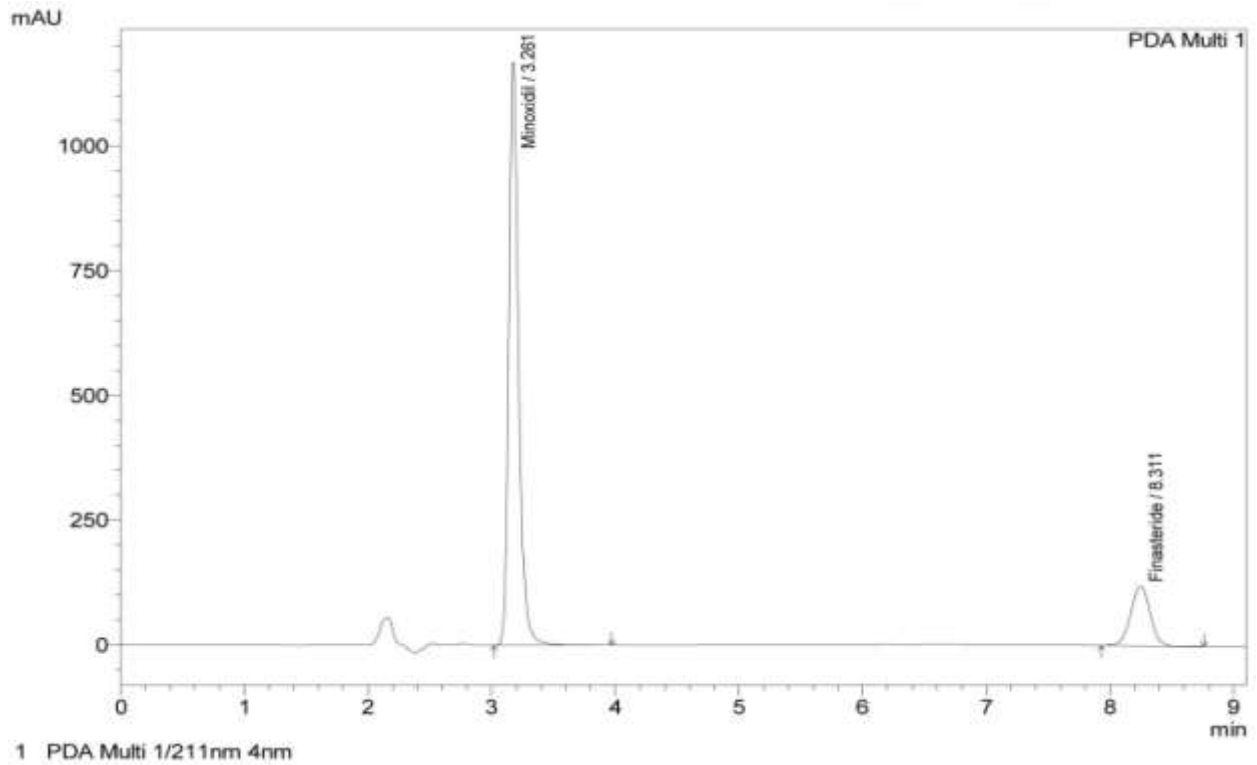


Figure 3 Sample chromatogram